# Choline and DHA supplementation ameliorate hippocampal damage in prenatally stressed rats by reducing apoptosis and cortisol levels

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Abstract. Aim: Prenatal stress inhibits neurogenesis and increases apoptosis in the hippocampus. Both Choline [C] and docosahexaenoic acid [DHA] are essential nutrients, important for the formation of neural cell membrane phospholipid bilayer. The neuroprotective potential of synergistic supplementation of these nutrients on the outcome of hippocampal neural cell density and neural development in prenatally stressed gestation is seldom evaluated. This study assesses the role of supplemented choline and or DHA in prenatally stressed neonates on their hippocampal neural cell density and serum cortisol levels. Materials and Methods: Pregnant rat dams were separated into [NC] - Normal control, [SC] - Saline control, [STR] - Stress, [STR+C] - Stress+Choline, [STR+DHA] - Stress+DHA, and [STR+C+DHA] -Stress +Choline+DHA groups [n= 6/group]. NC dams were undisturbed throughout the gestation. All other experimental groups of dams were supplemented by Saline, C, and DHA throughout the gestation respectively. All STR group dams were exposed with restraint stress from E11 till delivery. On postnatal day 40, pups were sacrificed after blood sample collection to estimate cortisol levels. Results: Cresyl violet stained, and caspase-3 labeled hippocampal sections were observed to analysis neural cell density and apoptosis. Significant restoration (p<0.001) in the total number of viable neuronal cells in CA1and CA3 subregions of the hippocampus and reduced caspase-3 labeled apoptotic cells were observed in STR+C+DHA rat pups compared to the age-matched NC, SC and stressed pups. In addition, a significant reduction in (p<0.01) serum cortisol concentration was found in [STR+C+DHA] pups when compared with age-matched stressed rat pups. Conclusion: Oral supplementation of nutrition like choline and DHA during stress among rat dams restores stress induced neonate hippocampal neural cell density and diminishes apoptotic cells in CA1 and CA3 regions with reductions in serum cortisol levels.

**Key words:** apoptosis, cortisol, choline, DHA, hippocampal neural density, hippocampal development, prenatal stress, neuroprotection, oxidative stress, maternal supplementation

# Introduction

Human brain development includes neurulation, neuronal proliferation, neural migration, neural cell differentiation, myelination, and apoptosis that occurs in the third week of intrauterine life as a result of differentiation with the progenitor cells of the neurons and prolongs up to all over the lifetime (1) Hippocampus is a small part of the human and animal brain that lies in the temporal lobe of the cerebrum. It forms the

posterior part of the limbic lobe and is responsible for learning, and memory (2). In both animals and humans, prenatal stress has detrimental effects on the neurobiological and hormonal development of offspring, which can lead to abnormal changes in cognition and behavioral outcomes in adolescent (3,4). Exposure to prenatal stress in monkeys' results in decreased neurogenesis in hippocampus and dentate gyrus (DG), leading to a decrease in the hippocampal volume (5). Exposure to abnormal levels of cortisol in the fetus causes a reduction in the number of neurons in hippocampus that results in reduction of hippocampal size (6). Prenatal stress induces a reduction in hippocampal volume and reduces postnatal neurogenesis in experimental animals. This decrease in hippocampal volumes/weight is due to the reduced number of hippocampal neural cells in prenatally stressed rats compared to normal control rat pups (7). Other animal studies also report that prenatal stress leads to depletion in neuron proliferation in hippocampus(8,9). Prenatal stress causes premature birth, low birth weight, fearfulness, and an increased risk of respiratory and skin illnesses in life (10). Another study reported that maternal stress affects heart rate of the fetus, intrauterine activity of the fetus, sleep cycle, leads to depression, and anxiety in postnatal life (11). Many human studies have expressed that pregnant woman who exposed to stress due to poor economic status, loss of relatives leads to sever consequences to their offspring such as autism and schizophrenia (12). Hormone cortisol plays a crucial role during stress in humans, monkeys, and rodents and affects the hypothalamic-pituitary-adrenal (HPA) axis (13). Chronic prenatal and/or early postnatal stress, resulting in dysregulation of negative HPA feedback via altered glucocorticoid receptor sensitivity to glucocorticoids (14). Stress causes the secretion of high levels of glucocorticoids which decreases glucocorticoid receptors in the hippocampus. The above findings clearly indicate that the negative feedback by HPA axis is impaired. Various human studies show that hyperactivity of the HPA axis is linked with low birth weight (15,16). The higher and prolonged glucocorticoid exposure affects neurotransmitter systems in the brain, especially hippocampus, serotonergic, dopaminergic, GABA-ergic, and noradrenergic systems (17-19). Calcium supplementation in pregnancy

also plays a role in reducing maternal and neonatal morbi-mortality by directly reducing the likelihood of developing pre-eclampsia and eclampsia (20). Variations of blood ion concentrations in pregnancy do not only affect maternal physiology but go a long way to impact fetal outcomes (21). Choline is one of the precursors for compounds, like phospholipids, acetylcholine, betaine, and is crucial for the developing fetus and neonate. Oral supplementation of choline for the pregnant mother and thereafter for the infant causes brain development which leads to changes to brain function (22). A high choline diet in pregnancy in animals increases hippocampal stem cell proliferation and inhibits apoptosis (23,24). Thus, in animals, prenatal choline supplementation enhances the brain structure and function of their offspring. Increased brain choline mediates memory function by elevating acetylcholine release (25). DHA enhances memory function by acting on the synaptic membrane fluidity cell signaling and regulating the gene expression (26). The availability of DHA promotes the differentiation of stem cells of central nervous system. In addition, DHA converts these stem cells into mature neural cells, thereby DHA influencing brain development (27). Animals supplemented with DHA during fetal life have high brain DHA which increases hippocampal neurogenesis. In vitro investigation also shows the efficiency of DHA in promoting neurogenesis. DHA not only facilitates the proliferation of embryonic stem cells but also causes neurite outgrowth of differentiated neural cells (28). DHA is vital for neurogenesis and neuritogenesis in prenatal and postnatal development (29). Until recently, no specific therapy has been developed to overcome stress-related neurodevelopmental deficits of the offspring, from dams exposed to stress during gestation. The first three months of pregnancy would be critical since medication would cause malformations in fetus. It is also recommended that certain medications are not to be used by pregnant women, even though no harmful effects have been observed (30). By considering all the above facts, this study was carriedout to analyze the efficacy of individual or combined supplementation of C and DHA during gestational stress in dams on the outcome of hippocampal neural cell density as well as serum cortisol levels in their neonates.

#### Materials and Methods

In-house bred albino Wistar strain adult male and female rats, obtained from the Central animal research facility, Manipal University, and their neonatal male and female rat pups were used in this study. All pups were maintained at 12:12 hrs. day: night environment, in a well-ventilated room in the Central animal research facility. Rats were fed with water and food *ad libitum*. Experiments were conducted only after the approval from institutional animal ethical committee with the approval number (IAEC/KMC/32/2012) that obey the guidelines enacted by the CPCSEA, New Delhi [India]. Proper care was taken while humanely handling the rats and all precautions were made and, we have used minimum number of animals for generating the necessary data.

# Experimental design

E0 day pregnant rat dams were divided into following groups: NC, SC, STR, STR+C, STR+DHA, and STR+C+DHA. Dams from NC group were undisturbed throughout the gestation period. Dams of SC group were supplemented orally with saline for the entire gestation period. STR group of dams were subjected to restraint stress from E11 to delivery. Pregnant dams from the STR+C group were supplemented with Choline from E0 to delivery and subjected to restraint stress from E11 until delivery. Dams from STR+DHA were supplemented with DHA from E0 until delivery and subjected to restraint stress from E11 until delivery. Dams of the STR+C+DHA group were supplemented orally with Choline and DHA from E0 until delivery and subjected to restraint stress from E11 to until delivery. Choline [Extra Pure choline chloride 98% was obtained from Loba Chemical Laboratory Reagents and Fine Chemicals] and dissolved with distilled water to make a dosage of (4.6 mmol/kg/day of choline) (31), and DHA [gelatin capsules consisting of 300 mg DHA were procured from Nouveau Medicament (P) Ltd., located at Chennai] (400 mg/day of DHA) (32). was supplemented orally to the dams using feeding needles. The need for choline and DHA during pregnancy is very high and the demand for these nutrients is increased workload by maternal organs, and

to support exponential fetal organ growth (33). Until recently, however, no specific drug therapies have been developed, which is administered during pregnancy, especially when the mother is in the stressful situation, would enhance the development of neurons in CNS especially in the hippocampus of offspring. The first 12 weeks of pregnancy is a critical time when medication can cause malformations of the fetus. So, experts may advise discontinuing medicines over this period unless a woman had multiple episodes of severe stress and depression. It is recommended that certain medications are not used by pregnant women, even though no harmful effects have been observed. The primary dietary factors involved in maintaining homeostasis and energy requirements such as calorie restriction, lipids, vitamins and other special nutritional supplements such as choline and DHA (30). Blood samples from each of the pups were collected for the estimation of cortisol. Then the pups were sacrificed after transcardial perfusion with saline and 10% formalin on postnatal day 40. Their brains were removed without any damage and processed for paraffin sectioning. Altogether, 300 sections of 5  $\mu$  thickness were made from the hippocampus. One section of every 30 sections were processed for cresyl violet staining. Randomly selected viable neural cells (cell body of the neurons) from 250  $\mu$ m area of CA1, CA3, and CA4 regions of the hippocampus, as also from the upper blade portion of the dentate gyrus (DG), were quantified using an ocular micrometer scale by another experimenter, blinded to the study. Non-viable or degenerated neurons were darkly stained, shrunken with fragmented nuclei were excluded from the count. Blinding of the slides from different groups were done prior to the counting to avoid observer bias. Photomicrographs were recorded under the compound Olympus microscope using cellSens Imaging Software at 40x magnification. The apoptosis of hippocampal neurons of CA1 and CA2 regions were observed by labeling caspase-3 expression using the immunofluorescence technique. Immunohistochemistry technique for estimating the apoptosis marker caspase-3 is based on the principle of antigen-antibody interactions. The hippocampal subregions were targeted with rabbit anti-caspase-3 primary antibodies which bind to express caspase-3 enzymes. After appropriate processing, the secondary

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sheep antibody tagged with CY-3 immunofluorescence were targeted to bind the primary antibody, and the fluorescence was identified and photomicrographed with a confocal microscope.

#### Restraint stress procedure

Restraint stress was performed from embryonic day 11 until delivery. The stress protocol involved placing the pregnant female in a wire mesh restrainer 6 hours per day. Control dams were left undisturbed throughout gestation. The wire mesh restrainer has a wooden base and stainless-steel wire mesh restrainer hinged to the base. A padlock and latch will help to secure the rat in the restrainer. The restrainers of two different dimensions used. The restrainer with 11 cm (length)  $\times$  6 cm (breadth)  $\times$ 6 cm (height) dimensions for restraining the pregnant rats from E1-E17 and restrainer with 11 cm  $(length) \times 8$  cm  $(breadth) \times 8$  cm (height) dimensions will be used to stress the pregnant rats from E18 till delivery. This type of restrainer claimed to restrict the animal's movement without any pain, discomfort or suffocation (34).

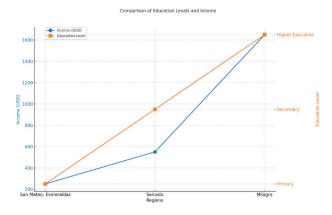
#### Statistical analysis

Data were analyzed using the method of one-way ANOVA and by Bonferroni's test and was expressed as mean  $\pm$  SEM. p < 0.05 was considered as significant. Entire analysis was done in software Graph pad prism version 5.03.

### Results

#### Serum cortisol levels in prenatally stressed rat pups

Rat pups subject to prenatal stress marked a significant increase (p<0.01) in mean serum cortisol level when compared with age-matched NC and SC group of rat pups. Rat pups subjected to prenatal stress and prenatal supplementation of choline marked a significant decrease (p<0.01) in the serum cortisol concentration when compared with the agematched rat pup groups which are exposed to prenatal stress (Figure 1).



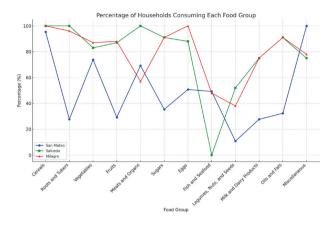
**Figure 1.** Mean serum cortisol level (ng/mL). n = 6 rats / group. NC & SC vs. STR [<sup>\*\*</sup>P<0.01], STR vs. STR+C+DHA [<sup>cc</sup>P<0.01], (One way ANOVA, Bonferroni's test). Normal control (NC), Saline control (SC), Stress (STR), Stress + Choline (STR+C), Sress+DHA (STR+DHA) and Stress+Choline+Docosahexaenoic acid (STR+C+DHA) group of rat pups.

# Prenatal stress and neural cells in hippocampal CA1 region

Rat pups subjected to prenatal stress exhibit a significant decrease (P<0.001) in mean number of viable neurons in CA1 region of the hippocampus when compared to NC & SC group of rat pups. STR+C, STR+DHA groups of rat pups exhibit significant increase (p<0.05) in the mean number of viable neurons, respectively, when compared to STR group of pups. However, STR+C+DHA rat pups exhibit significantly higher mean number of viable neurons (p<0.001) when compared to STR rat pups (Figures 2 and 3).

#### Prenatal stress and neural cells in hippocampal CA3 region

Rat pups subjected to prenatal stress exhibit significant decrease (P<0.001) in mean number of viable neurons in CA3 region when compared to NC & SC groups. Rat pups are subjected to prenatal stress with the supplementation of choline (p<0.05) or DHA (p<0.05) exhibit higher number of neurons,



**Figure 2.** Mean number of viable neural cells across 250μ length in CA1region of hippocampus; Mean ± SD. NC & SC vs. STR [\*\*\*P<0.001], STR vs. STR+C[<sup>a</sup>P<0.05], STR vs. STR+DHA[<sup>b</sup>P<0.05], STR vs. STR+C+DHA [<sup>ccc</sup>P<0.001], (One way ANOVA, Bonferroni's test) Normal control (NC), Saline control (SC), Stress (STR), Stress+Choline (STR+C), Stress+Docosahexaenoic acid (STR+DHA) and Stress+Choline+Docosahexaenoic acid (STR+C+DHA) group of rat pups.

respectively, when compared to STR groups. However, rat pups subjected to prenatal stress and undergone the supplementation of C+DHA exhibit significantly a greater number of neurons (p<0.001) when compared to STR group (Figures 4 and 5).

#### Prenatal stress and neural cells in hippocampal CA4 region

Rat pups exposed to prenatal stress exhibit significant (P<0.001) reduction in the mean number of viable neurons in the CA4 region when compared to NC & SC group of pups. Rat pups subjected to prenatal stress and undergone the supplementation of choline (p<0.05); DHA (p<0.05) and C+DHA exhibit a significantly greater (p<0.001) number of neurons when compared to STR group (Figures 6 and 7).

#### Prenatal stress and neural cells in hippocampal DG region

Rat pups exposed to prenatal stress showed significant reduction in the number of viable neurons (P<0.001) in DG region when compared to NC & SC groups. Rat pups subjected to prenatal stress and undergone the supplementation of choline (p<0.05); DHA (p<0.05) and C+DHA exhibit significantly

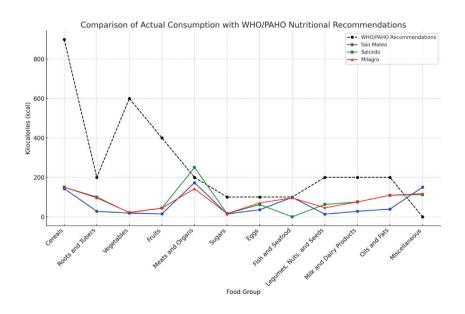
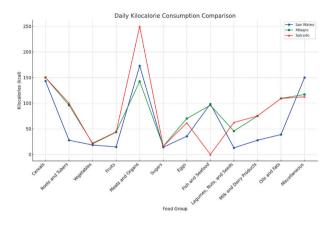


Figure 3. Photomicrographs of CA1 region of hippocampus. Normal control (NC), Saline control (SC), Stress (STR), Stress+Choline (STR+C), Stress+Docosahexaenoic acid (STR+DHA) and Stress+Choline+ Docosahexaenoic acid (STR+C+DHA) group of rat pups. (Cresyl violet stain X 10).



**Figure 4.** Mean number of viable neurons across 250µ length in CA3 region of hippocampus; Mean ± SD is shown. NC & SC vs. STR [\*\*\*P<0.001], STR vs. STR+C[<sup>a</sup>P<0.05], STR vs. STR+DHA[<sup>b</sup>P<0.05], STR vs. STR+C+DHA [<sup>ccc</sup>P<0.001], (One way ANOVA, Bonferroni's test) Normal control (NC), Saline control (SC), Stress (STR), Stress+Choline (STR+C), Stress+Docosahexaenoic acid (STR+DHA) and Stress+Choline+ Docosahexaenoic acid (STR+C+DHA) group of rat pups.

greater number of neurons (p<0.001) when compared with STR group of rat pups (Figures 8 and 9).

# Caspase – 3 immunofluorescence expression in neurons of hippocampal CA1 region

NC & SC group of rat pups showed very few and random signals of caspase - 3 in CA1 sub-region of hippocampus whereas the caspase - 3 expression of CA1 neurons of STR group were observed to be more while comparing to NC and SC groups. However caspase - 3 expressions of apoptotic CA1 neurons in hippocampal sections of all supplemented groups of rat pups [(STR+C), (Stress+DHA) and (STR+C+DHA)] were markedly reduced compared to STR group of rat pups (Figure 10).

# Caspase – 3 immunofluorescence expression in neural cells of hippocampal CA3 region

NC & SC group of rat pups showed very few and random signals of caspase - 3 in CA3 sub-region of hippocampus whereas the caspase - 3 expression of CA3 neurons of STR group were observed to be more while comparing to NC and SC groups. However

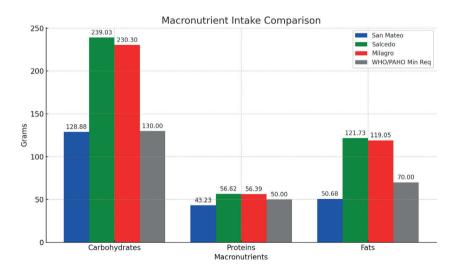
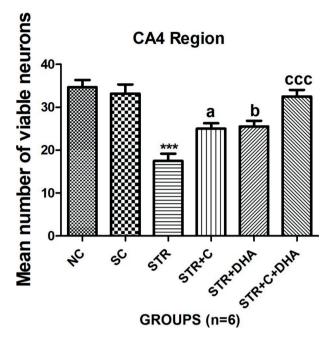


Figure 5. Photomicrographs of CA3 region of hippocampus. Normal control (NC), Saline control (SC), Stress (STR), Stress+Choline (STR+C), Stress+Docosahexaenoic acid (STR+DHA) and Stress+Choline+ Docosahexaenoic acid (STR+C+DHA) group of rat pups. (Cresyl violet stain X 10).



**Figure 6.** Mean number of viable neurons across 250µ length in CA4 region of hippocampus; Mean ± SD is shown. NC & SC vs. STR [\*\*\*P<0.001], STR vs. STR+C[<sup>a</sup>P<0.05], STR vs. STR+DHA[<sup>b</sup>P<0.05], STR vs. STR+C+DHA [<sup>ccc</sup>P<0.001], (One way ANOVA, Bonferroni's test) Normal control (NC), Saline control (SC), Stress (STR), Stress+Choline (STR+C), Stress+Docosahexaenoic acid (STR+DHA) and Stress+Choline+ Docosahexaenoic acid (STR+C+DHA) group of rat pups.

caspase - 3 expressions of apoptotic CA3 neurons in hippocampal sections of all supplemented groups of rat pups [(STR+C), (Stress+DHA) and (STR+C+DHA)] were markedly reduced compared to STR group of rat pups (Figure 11)

#### Discussion

Results from the present study shows that postnatal rat pups from gestationally stressed dams have significantly higher serum cortisol concentration/levels when compared with the rat pups from non-stressed age-matched NC and SC dams. High cortisol levels during pregnancy is associated with results in abortion of the fetus, delay in fetal growth, premature delivery, and very low birth weight of offspring, respectively (35,36). Constriction of placental arteries are one of the complications of the maternal stress, which leads to reduced blood flow to the fetus thereby decreasing the availability of essential nutrients and oxygen to the offspring (37). The amount of cortisol produces during stress will have an adverse effect on HPA axis of the developing fetus which suppresses fetal growth and cause premature delivery (38). Rat pups from gestationally

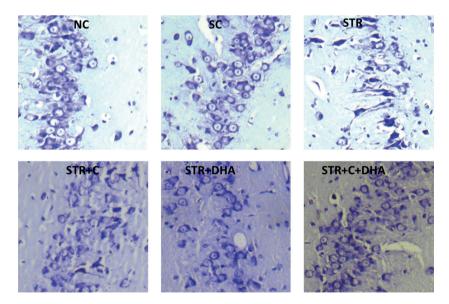
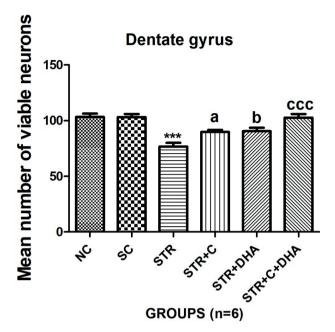
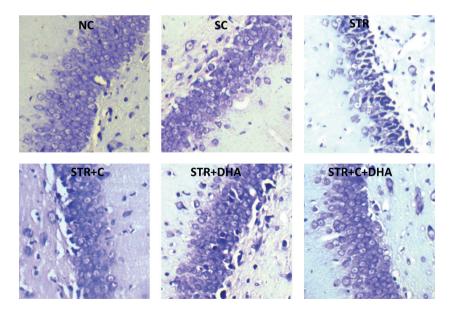


Figure 7. Photomicrographs of CA4 region of hippocampus. Normal control (NC), Saline control (SC), Stress (STR), Stress+Choline (STR+C), Stress+Docosahexaenoic acid (STR+DHA) and Stress+Choline+ Docosahexaenoic acid (STR+C+DHA) group of rat pups. (Cresyl violet stain X 10).

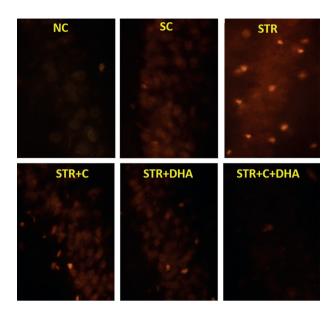


**Figure 8.** Mean number of viable neurons across 250 $\mu$  length in DG region of hippocampus; Mean ± SD is shown. NC & SC vs. STR [\*\*\*P<0.001], STR vs. STR+C[aP<0.05], STR vs. STR+DHA[<sup>b</sup>P<0.05], STR vs. STR+C+DHA [<sup>ccc</sup>P<0.001] (One way ANOVA, Bonferroni's test) Normal control (NC), Saline control (SC), Stress (STR), Stress+Choline (STR+C), Stress+Docosahexaenoic acid (STR+DHA) and Stress+Choline+ Docosahexaenoic acid (STR+C+DHA) group of rat pups.

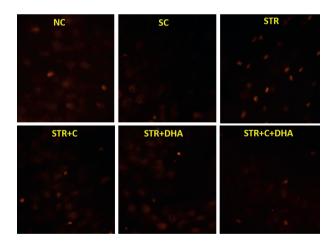
stressed dams when supplemented prenatally with choline and DHA showed a significant decrease in serum cortisol concentrations when compared to rat pups from age-matched non-supplemented stressed dams. These findings could be better explained by understanding various mechanisms through which choline produces higher methylation of CRH (corticotropin releasing hormone) genes of the placenta, this results in lower CRH transcription in the placenta, which leads to low concentrations of cortisol in the cord blood of the fetus. Studies have also reported that prenatal choline supplementation regulates the expression of genes that alter the fetal HPA axis sensitivity via epigenetic mechanisms (39,40). PUFAs, especially DHA and arachidonic acid, are fundamental elements of membrane phospholipids of the brain which are needed for optimal cerebral function (41). The hippocampus is the major constituent of the limbic system, which regulates the stress response. This part of the brain is also vulnerable to Glucocorticoids since mineralocorticoid receptors and glucocorticoid receptors are expressed in this limbic region (42). Feeding dams with DHA is proved to prevent stress-induced neural cell damage, apoptosis, and mitochondrial damages in hippocampus of their fetuses as we observed



**Figure 9.** Photomicrographs of DG region of hippocampus. Normal control (NC), Saline control (SC), Stress (STR), Stress+Choline (STR+C), Stress+Docosahexaenoic acid (STR+DHA) and Stress+Choline+ Docosahexaenoic acid (STR+C+DHA) group of rat pups. (Cresyl violet stain X 10).



**Figure 10.** Photomicrographs of CA1 neurons showing the signals of caspase -3 expression in different groups. Normal control (NC), Saline control (SC), Stress (STR), Stress + Choline (STR+C), Sress+DHA (STR+DHA) and Stress+Choline+Docosahexaenoic acid (STR+C+DHA) group of rat pups.



**Figure 11.** Photomicrographs of CA3 neurons showing the signals of caspase -3 expression in different groups. Normal control (NC), Saline control (SC), Stress (STR), Stress + Choline (STR+C), Sress+DHA (STR+DHA) and Stress+Choline+ Docosahexaenoic acid (STR+C+DHA) group of rat pups.

in our histochemical evaluations (43). Gestational stress affecting the fetus prenatally caused significantly lower numbers of surviving neural cells in postnatal rat hippocampus specifically in CA1, CA3, CA4, and DG regions when compared with NC and SC group

of rat pups. The number of surviving neurons in these sub-regions of the hippocampus was observed to be significantly increased by prenatal supplementation of choline and DHA alone or together during gestational stress. Additionally, surviving neural cells were observed to be significantly greater when both choline and DHA were supplemented in combination during the prenatal stress period. Chronic stress is associated with structural changes in the sub-regions of the hippocampus and can impair neurogenesis in the DG region (44). Maternal stress during gestation reduces hippocampal neurogenesis in primates (5). Maternal restraint stress even during the third trimester of gestation causes a significant decrease in neuronal proliferation in dentate gyrus (45). Maternal stress in mice reduces the number of dendritic spines and neuronal synapses in CA3 pyramidal cells (46). Neuronal proliferation of the hippocampus was decreased in PND 10 male rat pups in addition to the apoptosis in the sub-regions of the hippocampus caused by the chronic stress (47). Another study reported that stress causes elevation of cortisol and reductions in body weight and apoptosis in sub-regions of the hippocampus and cortex (48). Prenatal supplementation of choline, which is an important methyl donor, influences neurogenesis and apoptosis in fetal hippocampus (49). Apoptosis in the nervous system is modulated by neurotrophins and sex hormones (50). It has been proved that cholineregulated intermediate signals that mediate apoptosis are induced by deficiency of choline in the cortex and hippocampus of the fetus (51,52). Choline deficiency was associated with a decrease in phosphatidyl choline concentrations in some subcellular compartments, leading to the induction of apoptosis (53). Thus, choline deficiency causes apoptosis in neurons by altering the cell cycle, as these neurons normally do not progress past G0/G1 (52). Representative photomicrographs of CA1 and CA3 sub-regions of the hippocampus from NC and SC rat pups were observed to have very few and arbitrary signals of caspase - 3 expression. Whereas caspase - 3 expressions were observed to be more in CA1 and CA3 neurons from prenatally stressed rat pups when compared to nonstressed NC and SC rat pups. Moreover, caspase - 3 expression in CA1 and CA3 neurons of hippocampal regions were found to be markedly reduced in

prenatally stressed rats supplemented with choline and DHA separately or both together prenatally when compared to non-supplemented prenatally stressed rat pups. Prenatal supplementation of choline causes neurogenesis by preventing apoptosis in the developing hippocampus (49). Apoptosis is programmed cell suicide, appearing even during normal development (54). It has been proved that apoptosis in the developing hippocampus as well as other part of the brain is facilitated by choline deficiency (51). Choline deficiency associated with a decrease in PtdCho concentrations, and some subcellular compartments leads to the induction of apoptosis (55). Thus, choline deficiency causes apoptosis in neurons by the disturbance in the cell cycle, as a result, cell division is impaired (52). Maternal choline supplementation during E11 and 17 results in significant changes of the developing hippocampus and other parts of the brain by altering apoptosis (51). Availability of choline during development stimulates neuronal cell division of the brain whereas choline deficiency inhibits proliferation of neuronal precursor cells and stimulates apoptosis (23,55). Supplementation of choline in gestation boosts up neurogenesis in the hippocampus and dentate gyrus (56). This effect on adult neurogenesis is believed to be brought by increased hippocampal concentrations of trophic factors like BDNF, insulin like growth factor 2 (IGF2), as well as vascular endothelial growth factor (VEGF) which help to increase the size of cholinergic neurons of the brain, facilitate acetylcholine synthesis and better cognitive function (57). Recent studies explore the coordinated roles of metabolism of choline and DHA during fetal development (58). The metabolic coordination of DHA and C has been well explained in vivo, in Pemt-/- mouse model. The PEMT enzyme increases the synthesis of phosphatidylcholine by using phosphatidylethanolamine as a precursor, that increases the requirement of choline in the tissues. Maternal DHA supplementation increases the concentration of brain PEMT enzyme thereby it increases the production of choline (55). Additionally, PEMT prefers phosphatidylethanolamine that contains DHA which is the long-chain PUFA and increases the development of DHA-enriched phosphatidylcholine in cell membranes of the neurons mainly in the brain (59). This metabolic coordination between the choline and

DHA through the PEMT enzyme restores neural cell density in the hippocampus by reduced apoptosis and low serum cortisol concentration.

# Conclusion

In conclusion, our findings documenting for the first time that supplementation of choline-DHA during prenatal stress in rats restores stress-induced neonate hippocampal neural cell density in CA1, CA3, and CA4 subregions of the hippocampus, and the dentate gyrus and diminishes apoptotic cells in CA1 and CA3 regions with reductions in serum cortisol levels. The result of this study suggests for further studies to revel the mechanism and the implication of the findings in humans.

Acknowledgments: The authors are thankful to Kasturba Medical College Manipal and Manipal Academy of Higher Education for administrative permission and for providing infrastructure for the research study.

**Conflict of Interest:** Each author declares that he or she has no commercial associations (e.g. consultancies, stock ownership, equity interest, patent/licensing arrangement etc.) that might pose a conflict of interest in connection with the submitted article.

Author Contribution: KSR, KMRB and RHT were involved in the conception and design of the study. Data collection and analysis were performed by RHT and. The first draft of the manuscript was written by RHT, and it was critically reviewed by KSR. The final draft of the manuscript was approved by all the authors.

**Limitation:** Study was limited to early adolescent period. The study showed only the neural cell density and serum cortisol of particular age group of rat pops. The mechanisms for alterations in neural cell density and serum cortisol need to be evaluated.

# References

- 1. Stiles J, Jernigan TL. The Basics of Brain Development. Neuropsychology Review. 2010 Nov 3;20(4):327–48. doi: 10.1007/s11065-010-9148-4.
- 2. Dhikav V, Anand KS. Hippocampus in Health and disease: an Overview. Annals of Indian Academy of Neurology. 2012;15(4):239–46. doi: 10.4103/0972-2327.104323.

- Kofman O. The role of prenatal stress in the etiology of developmental behavioural disorders. Neuroscience & Biobehavioral Reviews. 2002 Jun;26(4):457–70. doi:10.1016 /S0149-7634(02)00015-5.
- Coussons, Read ME. Effects of Prenatal Stress on Pregnancy and Human development: Mechanisms and Pathways. Obstetric Medicine. 2013 May 3;6(2):527. doi: 10.1177 /1753495X12473751.
- Coe CL, Kramer M, Czéh B, et al. Prenatal stress diminishes neurogenesis in the dentate gyrus of juvenile Rhesus monkeys. Biological Psychiatry. 2003 Nov;54(10):1025–34. doi:10.1016/S0006-3223(03)00698-X.
- Avishai-Eliner S. Stressed-out, or in (utero)? Trends in Neurosciences. 2002 Oct 1;25(10):518–24. doi:10.1016 /S0166-2236(02)02241-5.
- T. Szuran, Zimmermann E, H. Welzl. Water maze performance and hippocampal weight of prenatally stressed rats. Behavioural Brain Research. 1994 Dec 1;65(2):153–5. doi: 10.1016/0166-4328(94)90100-7.
- Kawamura T, Chen J, Takahashi T, Ichitani Y, Nakahara D. Prenatal stress suppresses cell proliferation in the early developing brain. NeuroReport. 2006 Oct 2;17(14):1515–8. doi: 10.1097/01.wnr.0000236849.53682.6d.
- 9. Zuena AR, Mairesse J, Casolini P, et al. Prenatal restraint stress generates two distinct behavioral and neurochemical profiles in male and female rats. PLoS One. 2008 May 14;3(5):e2170. doi: 10.1371/journal.pone.0002170. PMID: 18478112; PMCID: PMC2366064.
- Beijers R, Jansen J, Riksen-Walraven M, de Weerth C. Maternal Prenatal Anxiety and Stress Predict Infant Illnesses and Health Complaints. PEDIATRICS. 2010 Jul 19; 126(2):e401–9. doi:10.1542/peds.2009-3226.
- Kinsella MT, Monk C. Impact of maternal stress, depression and anxiety on fetal neurobehavioral development. Clinical Obstetrics and Gynecology. 2009 Sep;52(3): 425–40. doi: 10.1097/GRF.0b013e3181b52df1.
- Kinney DK, Munir KM, Crowley DJ, Miller AM. Prenatal stress and risk for autism. Neuroscience and biobehavioral reviews. 2008 Oct 1;32(8):1519–32. doi: 10.1016/j.neubiorev .2008.06.004.
- O'Regan D, Welberg LLAM, Holmes MC, Seckl JR. Glucocorticoid programming of pituitary–adrenal function: mechanisms and physiological consequences. Seminars in Neonatology. 2001 Aug;6(4):319–29.doi:10.1053 /siny.2001.0067.
- Tomas C, Newton J, Watson S. A Review of Hypothalamic-Pituitary-Adrenal Axis Function in Chronic Fatigue Syndrome. ISRN Neuroscience. 2013;2013:1–8. doi:10.1155 /2013/784520.
- Phillips DIW, Barker DJP, Fall CHD, et al. Elevated Plasma Cortisol Concentrations: A Link between Low Birth Weight and the Insulin Resistance Syndrome?1. The Journal of Clinical Endocrinology & Metabolism. 1998 Mar 1;83(3): 757–60. doi:10.1210/jcem.83.3.4634.
- 16. Levitt N, Lambert EV, Woods D, Hales CN, Andrew R, Seckl JR. Impaired Glucose Tolerance and Elevated Blood

Pressure in Low Birth Weight, Nonobese, Young South African Adults: Early Programming of Cortisol Axis1. The Journal of Clinical Endocrinology and Metabolism. 2000 Dec 1;85(12):4611–8. doi:10.1210/jcem.85.12.7039.

- Schneider ML, Roughton EC, Koehler AJ, Lubach GR. Growth and development following prenatal stress exposure in primates: an examination of ontogenetic vulnerability. Child Development. 1999 Mar 1 [cited 2020 Apr 4]; 70(2):263–74 doi:10.1111/1467-8624.00020.
- 18. Huizink AC, Robles de Medina PG, Mulder EJH, Visser GHA, Buitelaar JK. Stress during pregnancy is associated with developmental outcome in infancy. Journal of Child Psychology and Psychiatry, and Allied Disciplines. 2003 Sep 1;44(6):810–8. doi:10.1111/1469-7610.00166.
- Sapolsky R, Uno H, Rebert C, Finch C. Hippocampal damage associated with prolonged glucocorticoid exposure in primates. The Journal of Neuroscience. 1990 Sep 1;10(9): 2897–902. doi: 10.1523/JNEUROSCI.10-09-02897.1990.
- 20. Atem Bethel Ajong, Kenfack B, Innocent Mbulli Ali, et al. Calcium supplementation in pregnancy: An analysis of potential determinants in an under-resourced setting. PLOS ONE. 2023 Oct 5;18(10):e0292303–3. doi:10.1371/journal .pone.0292303.
- Ajong AB, Yakum MN, Aljerf L, et al. Association of hypertension in pregnancy with serum electrolyte disorders in late pregnancy among Cameroonian women. Scientific Reports. 2023 Nov 28 [cited 2024 Jul 30];13:20940. doi:10.1038 /s41598-023-47623-6.
- 22. Zeisel SH. The fetal origins of memory: the role of dietary choline in optimal brain development. The Journal of pediatrics. 2006 Nov 1;149(5 Suppl):S131–6. doi:10.1016 /j.jpeds.2006.06.065.
- 23. Albright CD, Friedrich CB, Brown EC, Mar MH, Zeisel SH. Maternal dietary choline availability alters mitosis, apoptosis and the localization of TOAD-64 protein in the developing fetal rat septum. Developmental Brain Research. 1999 Jun;115(2):123–9. doi:10.1016/S0165-3806(99)00057-7.
- 24. Albright CD, Tsai AY, Friedrich CB, Mar MH, Zeisel SH. Choline availability alters embryonic development of the hippocampus and septum in the rat. Developmental Brain Research. 1999 Mar;113(1-2):13–20. doi:10.1016 /S0165-3806(98)00183-7.
- Thomas JD, O'Neill TM, Dominguez HD. Perinatal choline supplementation does not mitigate motor coordination deficits associated with neonatal alcohol exposure in rats. Neurotoxicology and Teratology. 2004 Feb 28;26(2):223–9. doi:10.1016/j.ntt.2003.10.005.
- 26. Wu A, Ying Z, Gomez-Pinilla F. Dietary Omega-3 Fatty Acids Normalize BDNF Levels, Reduce Oxidative Damage, and Counteract Learning Disability after Traumatic Brain Injury in Rats. Journal of Neurotrauma. 2004 Oct;21(10): 1457–67 doi:10.1089/neu.2004.21.1457.
- Kan I, Melamed E, Offen D, Green P. Docosahexaenoic acid and arachidonic acid are fundamental supplements for the induction of neuronal differentiation. Journal of Lipid Research. 2007 Mar;48(3):513–7. doi:10.1194/jlr.C600022-JLR200.

- 28. He C, Qu X, Cui L, Wang J, Kang JX. Improved spatial learning performance of fat-1 mice is associated with enhanced neurogenesis and neuritogenesis by docosahexaenoic acid. Proceedings of the National Academy of Sciences of the United States of America. 2009 Jul 7 [cited 2022 Dec 14];106(27):11370–5. doi:10.1073/pnas.0904835106.
- Tailby C, Wright LL, Metha AB, Calford MB. Activitydependent maintenance and growth of dendrites in adult cortex. Proceedings of the National Academy of Sciences. 2005 Mar 14;102(12):4631–6. doi:10.1073/pnas.0402747102.
- Koren G, Pastuszak A, Ito S. Drugs in Pregnancy. Wood AJJ, editor. New England Journal of Medicine. 1998 Apr 16;338(16):1128–37. doi: 10.1056/NEJM199804163381607.
- Zeisel SH. Choline: Needed for Normal Development of Memory. Journal of the American College of Nutrition. 2000 Oct;19(sup5):528S531S. doi:10.1080/07315724.200 0.10718976.
- 32. Sakamoto T, Cansev M, Wurtman RJ. Oral supplementation with docosahexaenoic acid and uridine-5'-monophosphate increases dendritic spine density in adult gerbil hippocampus. Brain Research. 2007 Nov;1182:50–9. doi: 10.1016 /j.brainres.2007.08.089.
- 33. Mun JG, Legette LL, Ikonte CJ, Mitmesser SH. Choline and DHA in Maternal and Infant Nutrition: Synergistic Implications in Brain and Eye Health. Nutrients. 2019 May 21;11(5):1125. doi:10.3390/nu11051125.
- 34. Sunanda null, Rao MS, Raju TR. Effect of chronic restraint stress on dendritic spines and excrescences of hippocampal CA3 pyramidal neurons–a quantitative study. Brain Research. 1995 Oct 2;694(1-2):312–7. doi:10.1016/0006-8993(95) 00822-8.
- 35. Campbell MK, Challis JRG, DaSilva O, Bocking AD. A cohort study found that white blood cell count and endocrine markers predicted preterm birth in symptomatic women. Journal of Clinical Epidemiology. 2005 Mar;58(3):304–10. doi: 10.1016/j.jclinepi.2004.06.015.
- 36. Field T, Diego M. Cortisol: The Culprit Prenatal Stress Variable. International Journal of Neuroscience. 2008 Jan;118(8):1181–205. doi:10.1080/00207450701820944.
- 37. Myers RE. Maternal psychological stress and fetal asphyxia: A study in the monkey. 1975 May 1;122(1):47–59. doi:10.1016 /0002-9378(75)90614-6.
- 38. Challis JRG, Sloboda D, Matthews SG, et al. The fetal placental hypothalamic–pituitary–adrenal (HPA) axis, parturition and post natal health. Molecular and Cellular Endocrinology. 2001 Dec;185(1-2):135–44. doi: 10.1016 /S0303-7207(01)00624-4.
- Weinstock M. The long-term behavioural consequences of prenatal stress. Neuroscience & Biobehavioral Reviews. 2008 Aug;32(6):1073–86. doi: 10.1016/j.neubiorev.2008.03.002.
- 40. Jiang X, Yan J, West AA, et al. Maternal choline intake alters the epigenetic state of fetal cortisol-regulating genes in humans. FASEB journal: official publication of the Federation of American Societies for Experimental Biology. 2012 Aug 1;26(8):3563–74. doi: 10.1096/fj.12-207894.

- Alessandri JM, Guesnet P, Vancassel S, et al. Polyunsaturated fatty acids in the central nervous system: evolution of concepts and nutritional implications throughout life. Reproduction Nutrition Development. 2004 Nov 1; 44(6):509–38. doi: 10.1051/rnd:2004063.
- 42. Hennebelle M, Champeil-Potokar G, Lavialle M, Vancassel S, Denis I. Omega-3 polyunsaturated fatty acids and chronic stress-induced modulations of glutamatergic neurotransmission in the hippocampus. Nutrition Reviews. 2014 Jan 13;72(2):99–112. doi: 10.1111/nure.12088.
- 43. Feng Z, Zou X, Jia H, et al. Maternal Docosahexaenoic Acid Feeding Protects Against Impairment of Learning and Memory and Oxidative Stress in Prenatally Stressed Rats: Possible Role of Neuronal Mitochondria Metabolism. Antioxidants & Redox Signaling. 2012 Feb 1;16(3):275–89. doi:10.1089/ars.2010.3750.
- 44. MagariñosAM, McEwen BS, Flügge G, Fuchs E. Chronic Psychosocial Stress Causes Apical Dendritic Atrophy of Hippocampal CA3 Pyramidal Neurons in Subordinate Tree Shrews. The Journal of Neuroscience. 1996 May 15;16(10):3534–40. doi: 10.1523/JNEUROSCI.16-10-03534.1996.
- 45. Fujioka A, Fujioka T, Ishida Y, Maekawa T, Nakamura S. Differential effects of prenatal stress on the morphological maturation of hippocampal neurons. Neuroscience. 2006;141(2):907–15. doi: 10.1016/j.neuroscience.2006.04.046.
- 46. Ishiwata H, Shiga T, Okado N. Selective serotonin reuptake inhibitor treatment of early postnatal mice reverses their prenatal stress-induced brain dysfunction. Neuroscience. 2005 Jan;133(4):893–901. doi: 10.1016/j.neuroscience.2005.03.048.
- 47. Joëls M, Karst H, Alfarez D, et al. Effects of Chronic Stress on Structure and Cell Function in Rat Hippocampus and Hypothalamus. Stress. 2004 Dec;7(4):221–31. doi: 10.1080/10253890500070005.
- 48. Lucassen PJ, Vollmann-Honsdorf GK, Gleisberg M, Czéh B, De Kloet ER, Fuchs E. Chronic psychosocial stress differentially affects apoptosis in hippocampal subregions and cortex of the adult tree shrew. European Journal of Neuroscience. 2001 Jul;14(1):161–6. doi: 10.1046/j.0953-816x.2001 .01629.x.
- 49. Craciunescu CN, Johnson AR, Zeisel SH. Dietary Choline Reverses Some, but Not All, Effects of Folate Deficiency on Neurogenesis and Apoptosis in Fetal Mouse Brain. The Journal of Nutrition. 2010 Jun 1;140(6):1162–6. doi:10.3945/jn.110.122044.
- Henderson CE. Programmed Cell Death in the Developing Nervous System. Neuron. 1996 Oct;17(4):579–85. doi: 10.1016/s0896-6273(00)80191-9.
- Holmes-McNary MQ, Loy R, Mar M-H, Albright CD, Zeisel SH. Apoptosis is induced by choline deficiency in fetal brain and in PC12 cells. Developmental Brain Research. 1997 Jul;101(1-2):9–16. doi: 10.1016/S0165-3806(97)00044-8.
- 52. Yen Cle, Mar MH, Meeker RB, Fernandes A, Zeisel SH. Choline deficiency induces apoptosis in primary cultures of fetal neurons. The FASEB Journal. 2001 Aug 1; 15(10):1704–10. doi: 10.1096/fj.00-0800com.

- 53. Shin OH, Mar MH, Albright CD, Citarella MT, Costa KA da, Zeisel SH. Methyl-group donors cannot prevent apoptotic death of rat hepatocytes induced by choline-deficiency. Journal of Cellular Biochemistry. 1997 Feb 1;64(2):196–208. doi:10.1002/(SICI)1097-4644(199702)64:2<196::AID-JCB3>3.0.CO;2-S.
- Schwartzman RA, Cidlowski JA. Apoptosis: The Biochemistry and Molecular Biology of Programmed Cell Death\*. Endocrine Reviews. 1993 Apr;14(2):133–51. doi: 10.1210/ edrv-14-2-133.
- 55. Ann K, Rai KS, Corneliu Marius Cra ciunescu, Parikh K, et al. Dietary Docosahexaenoic Acid Supplementation Modulates Hippocampal Development in the Pemt-/- Mouse. 2010 Jan 8;285(2):1008–15. doi:10.1074/jbc.M109.017137.
- 56. Gonçalves JT, Schafer ST, Gage FH. Adult Neurogenesis in the Hippocampus: From Stem Cells to Behavior. Cell. 2016 Nov;167(4):897–914. doi:10.1016/j.cell.2016 .10.021.
- 57. Napoli I, Blusztajn JK, Mellott TJ. Prenatal choline supplementation in rats increases the expression of IGF2 and its receptor IGF2R and enhances IGF2-induced acetylcholine release in hippocampus and frontal cortex. Brain Research. 2008 Oct;1237:124–35. doi: 10.1016/j.brainres.2008.08.046.

- 58. Thomas Rajarethnem H, Megur Ramakrishna Bhat K, Jc M, Kumar Gopalkrishnan S, Mugundhu Gopalram RB, Rai KS. Combined Supplementation of Choline and Docosahexaenoic Acid during Pregnancy Enhances Neurodevelopment of Fetal Hippocampus. Neurology Research International. 2017;2017:8748706. doi: 10.1155/2017/8748706.
- Ridgway ND, Vance DE. [43] Phosphatidylethanolamine N-methyltransferase from rat liver. Methods in enzymology on CD-ROM/Methods in enzymology. 1992 Jan 1;366–74. doi: 10.1016/0076-6879(92)09045-5.

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- Accepted: 9 December 2024
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