ORIGINAL ARTICLE

Novel micronutrient associations in homocysteine metabolism

Hari Krishnan Krishnamurthy¹, Swarnkumar Reddy, Vasanth Jayaraman¹, Karthik Krishna¹, Qi Song², Karenah E. Rajasekaran², Tianhao Wang¹, Kang Bei¹, John J. Rajasekaran¹

¹Vibrant Sciences LLC., San Carlos, CA, United States of America; ²Vibrant America LLC., San Carlos, CA, United States of America

Abstract. Objective: Elevated serum level of homocysteine is one of the most effective biomarker in diagnosing atherosclerotic cardiovascular disease and associated disorders. The elevated levels of homocysteine may result from mutations in genes coding vital enzymes or other environmental factors such as nutrition, medications, increased methionine intake, pregnancy, etc. The association of 36 vital micronutrients with circulating levels of homocysteine in a population of 336 free-living individuals was analyzed. Methods: Baseline serum homocysteine concentration was measured by enzyme-linked immunosorbent assay using Beckman Counter AU series Analyzers. Micronutrients were measured using LC-MS/MS Waters TQ-XS Tandem mass spectrometer. Results: Non-parametric analysis by Mann-Whitney U test as well as Pearson's correlation showed a strong association of vitamin B_9 (folate) and vitamin B_{12} with high serum levels of homocysteine. Interestingly several amino acids including asparagine, isoleucine, cysteine, citrulline, and carnitine exhibited a positive correlation with serum homocysteine. Among vitamins and minerals, vitamin A and selenium were positively correlated with circulating homocysteine. Conclusion: The study suggests that the serum levels of several micronutrients act as confounding variables in regulating the circulating levels of homocysteine. Several novel nutrient associations for modulating levels of homocysteine are highlighted. Given that homocysteine is a modifiable risk factor, these nutrients could play an important role in preventive care settings and should be further investigated clinically.

Key words: homocysteine, micronutrients, methionine, vitamins, amino acids, and cardiovascular diseases

Introduction

Cardiovascular disease (CVD) accounts for more than 25% of deaths in the United States and is also the major cause of morbidity. Diet and nutrition play a crucial role in the pathogenesis of CVD and are important factors that can be used to prevent or delay the onset of CVD (1-3). Predictive risk factors such as lipid concentrations, diabetes mellitus, and blood pressure are frequently used to measure the effect of diet on disease prevention (4). Studying these predictive biomarkers often measures a single factor, whereas

the effect of dietary changes involves multiple factors. Several observational studies have reported elevated levels of circulating homocysteine and its association with an increased risk of CVD. It is also an independent marker for atherosclerotic cardiovascular disease. Elevated levels of homocysteine are associated with various other metabolic disorders such as Alzheimer's disease and Parkinson's disease (5, 6). McCully postulated the theory of homocysteine-derived atherosclerosis in 1969. He had observed 2 rare distinct genetic conditions, a disorder in vitamin B₁₂ which inhibits the conversion of homocysteine to methionine in a vitamin

 B_{12} -dependent reaction, and cystathionine β -synthase deficiency might result in homocystinuria, cystathioninuria, and methylmalonic aciduria (7).

Homocysteine is a sulfhydryl-containing essential amino acid and an intermediate metabolite in the biosynthesis of cysteine from methionine. Dietary methionine is the most common source of homocysteine which is produced via the demethylation of methionine. The circulating levels of homocysteine are regulated via various micronutrient-dependent pathways (8, 9). Free homocysteine can be converted to methionine by re-methylation of circulating homocysteine, this reaction is catalyzed by enzymes N5, N10-methyltetrahydrofolate reductase as a methyl donor with cobalamin (vitamin B_{12}) as a cofactor (10). Cystine, a semi-essential amino acid is synthesized from free homocysteine catalyzed by the enzyme cystathionine β -synthase and pyridoxine (Vit B_6) as a cofactor (11).

Numerous case-control, retrospective, and prospective studies have reported the correlation between various essential micronutrients and serum homocysteine (12-14). The elevated levels of serum homocysteine may result from mutations in genes coding vital enzymes of homocysteine metabolism or other environmental factors such as increased methionine intake, medications, pregnancy, etc. In addition to these factors' deficiency of vitamins and vitamin, cofactors are the most preventable cause of hyperhomocysteinemia. Several reports have shown that supplementing certain vitamins such as folate, vitamin B12, and B6 reduces serum levels of homocysteine (15, 16). An imbalance of methionine, which is mostly found in animal proteins, and a lack of vitamins B6, B12, and folate, may result in increasing the level of homocysteine in the body. Reports have suggested that higher consumption of total folate was linked to a decreased risk of glaucoma, but the protective effect was more significant when folate was obtained through supplementation in addition to dietary intake (17, 18). All those studies widely discuss only vitamins, the present study attempted to demonstrate the clinical importance of the diverse classes of micronutrients. Various micronutrients and amino acids such as folate, vitamin B₁₂. and homocysteine are known to interact in one-carbon metabolism and are considered vital elements for cellular growth and differentiation (18). Hence, we hypothesized that the serum levels of micronutrients play

a crucial role in regulating free homocysteine, and in the present study we investigated the association of 36 vital micronutrients with circulating levels of homocysteine in a population of 336 free-living individuals.

Materials and methods

Study population

The study included a total of 336 free-living general population who were tested for cardiovascular panel and micronutrient panel at Vibrant America Clinical Laboratory with no clinical indications of abnormalities. This was a retrospective study on deidentified individuals and hence exempted from formal ethical review by Western IRB (IRB # 1-1288754-1) (Washington USA).

Determination of serum micronutrients and homocysteine concentrations

Blood samples were processed for the separation of serum and further analyzed for serum concentrations of various micronutrients and homocysteine. Total serum homocysteine levels were estimated by invitro quantitative enzyme-linked immunosorbent assay using Beckman Counter AU series Analyzers. The serum level of homocysteine is measured as the inverse of the amount of NADH converted to NAD+ which can be measured at 340nm. Serum levels of micronutrients were determined using LC-MS/MS Waters TQ-XS Tandem mass spectrometer, Waters GC-MS, and Perkin Elmer NexION ICP-MS instrumentation using standard protocols.

Statistical analysis

Retrospective analysis of clinical data from deidentified subjects was performed using Java for Windows version 1.8.161. Homocysteine serum levels were stratified into four quartiles (Q1 to Q4) based on the cutoff points at 25th, 50th, and 75th percentile values. The data was analyzed by a non-parametric Mann-Whitney U test for the evaluation of the association between serum levels of homocysteine and micronutrients. The univariant relationship between

serum levels of homocysteine and micronutrients was analyzed by Pearson's correlation with significance set at *p*<0.05. GraphPad Prism Version 7.00 was used to perform statistical analysis and descriptive statistics were used to express the continuous variables (mean ± SD, median, minimum, and maximum)

Results

The study comprised 336 individuals with a mean age of 48.7 ± 15.0 and mean homocysteine was 9.9 ± 3.0 µmol/L. The experimental population was stratified into quartiles with mean homocysteine of 6.9 ± 0.9 ,

 8.6 ± 0.3 , 10.2 ± 0.6 , and 14.0 ± 3.1 for 25^{th} , 50^{th} , 75^{th} , and >75th percentile values respectively. Age was found to have a positive trend and a strong significance with increasing levels of serum homocysteine (Table S1). Serum levels of cysteine and citrulline were also found to have a positive trend with an increase in serum levels of homocysteine. Vitamin B_9 (folate) and vitamin B_{12} were found to have strong significance and negative trends with increasing concentrations of homocysteine.

Non-parametric analysis by the Mann-Whitney U test showed a strong association between high serum levels of homocysteine with vitamin B_9 (folate) and vitamin B_{12} (Table 1). This result was further supported by Person's correlation analysis where vitamin

Table 1. Significant association of serum homocysteine with micronutrients.

		Greater than Reference Range (n=22)	Within range (n=314)	P
		Baseline (X±SD)	Baseline (X±SD)	(P<0.05)
	Vitamin B ₁	17.2±9.4	21.8±15.3	0.2445
Water-soluble vitamins	Vitamin B ₂	23.9±22.4	28.5±25.1	0.0682
	Vitamin B ₃	18.5±6.9	19.1±8.7	0.8930
ole v	Vitamin B ₅	87.4±97.2	109.6±87.9	0.0635
soluł	Vitamin B ₆	13.8±14.2	16.5±14	0.1386
ter-	Vitamin B ₉	9.6±4.7	13.9±3.9	<0.0001
Wa	Vitamin B ₁₂	566.3±259.9	747.5±330.8	0.0032
>	Vitamin C	0.44±0.28	0.52±0.52	0.8036
ns	Vitamin A	81.0±30.8	77.1±23.9	0.8206
ami	Vitamin D ₃	0.96±0.38	0.92±0.27	0.9322
e vit	Vitamin D ₂₅ OH	43.6±20.6	48.2±23.2	0.3695
lubl	Vitamin K ₁	1.5±1.0	1.5±1.4	0.7364
Fat-soluble vitamins	Vitamin K ₂	0.66±0.84	0.72±1.2	0.5161
	Vitamin E	13.7±5.3	13.4±4.6	0.7837
	Selenium	147 ± 19.0	143.2±29.6	0.1309
	Sodium	140.3±3.0	141.7±3.4	0.0560
	Potassium	4.4±0.43	4.5±0.39	0.2598
	Calcium	9.7±0.3	9.6±0.4	0.0580
Minerals	Manganese	0.72±0.47	0.82±0.38	0.0841
Min	Zinc	0.68±0.12	0.71±0.16	0.2706
I	Copper	1.0±0.3	1.0±0.3	0.2375
	Chromium	0.22±0.12	0.25±0.16	0.8063
	Iron	106.1±40.2	103.1±39.1	0.5325
	Magnesium	2.1±0.25	2.2±0.1	0.0855

		Greater than Reference Range (n=22)	Within range (n=314)	P
		Baseline (X±SD)	Baseline (X±SD)	(P<0.05)
rial	Asparagine	57.6 ± 10.9	53.5±12	0.0642
Non- essential AA	Serine	145.4 ± 32.7	147.8±34.4	0.8101
A	Isoleucine	77.7±26.4	68.4±20.1	0.1392
BCAA	Valine	255.3±55.9	245.5±55.8	0.3863
B	Leucine	170.8±34.0	163.3±34.9	0.3140
ully A	Cysteine	22.2 ± 14.6	18.1±8.8	0.4027
Conditionally essential AA	Glutamine	500 ± 103.1	504.8 ± 80.2	0.8700
ndit	Citrulline	32.3±13.2	30.0±8.1	0.9183
C)	Arginine	131.9±53.7	122.4±31.7	0.8612
	Coenzyme Q10	2.2±1.0	2.1±0.9	0.5241
	Choline	14±4.6	14.0±5.7	0.8070
	Inositol	36.5±9.3	37.1±10.1	0.9253
	Carnitine	30.0±6.0	27±7.2	0.0203
	Methylmalonic acid	0.3±0.11	0.3±0.17	0.5839

Baseline values are expressed as mean (X) ± standard deviation (SD). For intergroup p-value, an unpaired t-student test was used.

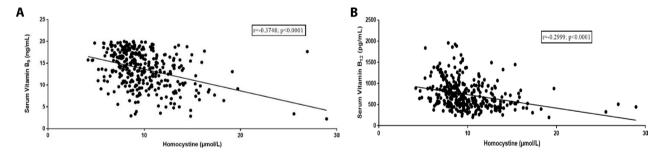


Figure 1. Pearson's correlation coefficients between homocysteine and micronutrients. (a) vitamin B₂. (b) vitamin B₁₂.

B₉ (r=-0.3748, p<0.0001) (Figure 1) and vitamin B₁₂ (r=-0.2999, p<0.0001) (Figure 2) exhibited a strong negative correlation with serum concentrations of homocysteine (Table 2). Vitamin A was found to have a significant positive correlation with homocysteine (r=0.1724, p<0.0015). Among the minerals analyzed selenium (r=0.1255, p<0.0214) was found to have a positive correlation with circulating homocysteine. In the case of amino acids, BCAA isoleucine, and nonessential amino acid asparagine were found to have a positive correlation with serum homocysteine. Among other amino acids, Cystine was found to have a strong

positive correlation (r=0.3216, p<0.0001) (Fig 3) whereas citrulline had a significant positive correlation with homocysteine.

Discussion

Homocysteine metabolism is regulated by several pathways depending on various metabolic enzymes and cofactors. Cystathionine β -synthase (CBS), methyltetrahydrofolate reductase (MTHFR), 5-methyltetrahydrofolate-homocysteine methyltransferase

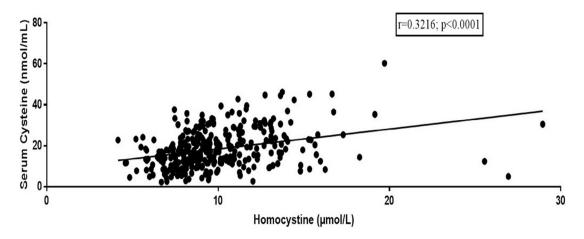


Figure 2. Pearson's correlation coefficients of homocysteine with cystine.

Table 2. Correlation of micronutrients with serum homocysteine.

		Correlation coefficient		
		r	p	
	Vitamin B ₁	-0.05311	0.3318	
ins	Vitamin B ₂	-0.07121	0.1929	
itam	Vitamin B ₃	0.0127	0.8165	
ole v	Vitamin B ₅	-0.09271	0.0898	
Water-soluble vitamins	Vitamin B ₆	-0.03308	0.5457	
ter-s	Vitamin B ₉	-0.3748	<0.0001	
Wa	Vitamin B ₁₂	-0.2999	<0.0001	
	Vitamin C	0.04608	0.3999	
su	Vitamin A	0.1724	0.0015	
ami	Vitamin D ₃	-0.00537	0.9219	
Fat-soluble vitamins	Vitamin D ₂₅ OH	-0.06897	0.2073	
lubl	Vitamin K ₁	0.02252	0.6809	
lt-so	Vitamin K ₂	-0.01648	0.7635	
L F	Vitamin E	-0.04578	0.4029	
	Selenium	0.1255	0.0214	
	Sodium	0.0395	0.4705	
	Potassium	-0.06882	0.2083	
	Calcium	0.05303	0.3325	
Minerals	Manganese	0.007346	0.8933	
Min	Zinc	-0.08034	0.1417	
	Copper	-0.05889	0.2818	
	Chromium	-0.0265	0.6283	
	Iron	0.004576	0.9334	
	Magnesium	0.03044	0.5781	

		Correlation coefficient		
		r	p	
al	Asparagine	0.1087	0.0464	
Non- essential AA	Serine	-0.01411	0.7967	
	Isoleucine	0.1326	0.015	
BCAA	Valine	0.04892	0.3714	
BC	Leucine	0.07977	0.1445	
Illy A	Cysteine	0.3216	<0.0001	
Conditionally essential AA	Glutamine	0.05482	0.3164	
ndit	Citrulline	0.1346	0.0135	
Co	Arginine	0.05025	0.3585	
	Coenzyme Q10	0.07233	0.186	
	Choline	0.08041	0.1413	
	Inositol	-0.01533	0.7795	
	Carnitine	0.1156	0.0342	
	Methylmalonic acid	0.09457	0.0835	

^{*}Data are given as Pearson's correlation (r) coefficients with adjustment for serum homocysteine

reductase (MTRR), methionine synthase (MTR), betaine-homocysteine S-methyltransferase (BHMT) are the major enzymes involved in homocysteine metabolism (19). Vitamin B_6 , vitamin B_9 (folate), and vitamin B_{12} are the vital cofactors for homocysteine metabolism. Upregulation of homocysteine can result

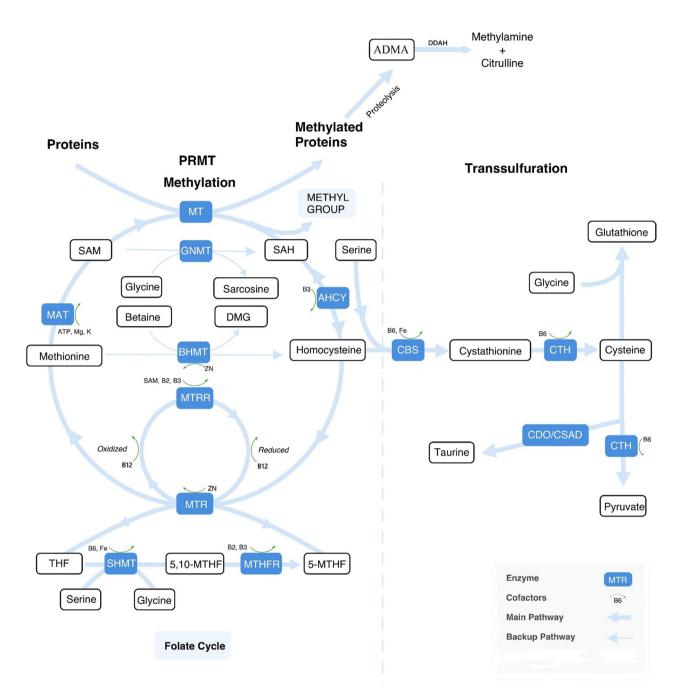


Figure 3. Diagrammatic representation of homocysteine metabolism detailing the interactions of cofactors vitamin B_6 , vitamin B_9 , and vitamin B_{12} . Abbreviations: PRMT: Protein Arginine Methyltransferase; ADMA: asymmetric dimethylarginine; MT: Methyl transferase; SAM: S-adenosyl methionine; SAH: S-adenosyl homocysteine; ACHY: Adenosylhomocysteinase; MAT: methionine adenosyltransferase; BHMT: Betaine-homocysteine methyltransferase; MTRR: methionine synthase reductase; SHMT: serine hydroxymethyltransferase; THF: Tetrahydrofolate; 5,10-MTHF: 5,10-methylene tetrahydrofolate; CBS: Cystathionine beta-synthase; CTH: cystathionine γ -lyase; CDO: Cysteine dioxygenase; CSAD: Cysteine Sulfinic Acid Decarboxylase; DDAH: Dimethylarginine Dimethylminohydrolase; DMG: Dimethylglycine; MTHFR: Methylenetetrahydrofolate reductase.

from two major factors, genetic enzymatic disorders, and inadequate cofactors (20). Several novel nutrient associations were found in our study.

Our population-based retrospective study of 336 individuals showed a linear relationship between serum levels of homocysteine with an increase in the age of the individuals and vital nutrients such as vitamin A, B₉, B₁₂, selenium, and amino acids such as asparagine, isoleucine, cysteine, and citrulline. These clinical variables particularly cysteine, vitamin B₉ (folate), and vitamin B_{12} are known to be regular harbingers of various metabolic disorders including chronic cardiovascular diseases while our study has found new associations due to the extensive testing of nutrients (21,22). Age is one of the prime causes of the upregulation of serum homocysteine resulting in various ageassociated disorders particularly poor wound healing, loss of regenerative ability, decreased renal and cognitive functioning, endothelial dysfunction, etc (23,24). Although the mechanism between the association of homocysteine with aging and associated disorders remains unclear, the current study clearly shows that the increase in the serum levels of homocysteine has a significant linear association with aging (Table 1). Ostrakhovitch et al. 2019 (23) have reported the association of elevated serum levels of homocysteine with aging. Age is associated with several other compilations such as reduction in absorption of proteins and other nutrients resulting in dietary restriction of vital vitamin cofactors such as vitamin B_6 , B_9 , and B_{12} these, in turn, lead to increased levels of serum homocysteine (25).

The inability to regulate the methionine pathway results in the accumulation of homocysteine which can be attributed to various endogenous and exogenous factors (26). The endogenous factors include SNPs in the genes coding the enzymes involved in methionine metabolism (Fig 3), which can be regulated by enzyme therapy or nutritional support (27). Nutritional support in the form of vital cofactors such as vitamin B_6 , B_9 , and B_{12} regulates the supply of methyl group and ensures the biochemical pathway for methylation of homocysteine to methionine or induces the transsulfuration pathway with the help of serine to convert

the serum homocysteine into cysteine (28). The current data demonstrated a negative trend between increasing serum levels of homocysteine with vitamins B₉ (folate) and B₁₂. Non-parametric analysis by Man Whitney and univariant analysis by Pearson's exhibited a significant association and a strong negative correlation of vitamins B₉ and B₁₂ with circulating levels of homocysteine. These results signify that a decrease in the serum level of vitamin B_9 and B_{12} increases the circulating levels of homocysteine by limiting the availability of methyl groups and inhibiting methylation. Numerous studies have reported the protective effect of vitamins by downregulating homocysteine. It's also been reported that dietary supplementation of vitamin B₁, vitamin B₂, vitamin B₉, and vitamin C was inversely associated with the development of hyperhomocysteinemia (29).

Our data exhibited a positive correlation between vitamin A and homocysteine. This association might result from the altered inflammatory and lipid metabolism at the hepatic level. Olsen et al. (2018) (30) observed a similar association where the patients with high total homocysteine were reported to have elevated levels of vitamin A. Inflammatory markers such as oxysterols produced during inflammatory conditions such as non-alcoholic fatty liver disease induces the hepatic expression of genes responsible for the synthesis of all-trans-Retinoic acid, a potential activator of glycine N-methyltransferase (GNMT). The activation of GNMT accelerates the synthesis of SAH (S-Adenosyl-L-homocysteine) and homocysteine (Fig 3) (31).

Minerals are the most vital nutrients with diverse functionalities in maintaining body homeostasis and metabolism. The present study included various vital minerals, among all the minerals selenium was noted with a positive correlation with serum homocysteine. Selenium is a vital trace mineral with wide interindividual variability in metabolic activities (32). Various reports have demonstrated selenium as a potential factor in altering total homocysteine. Considerable reports have described the role of selenoproteins in various metabolic pathways, particularly thyroid

synthesis, and homocysteine, but the detailed mechanism remains unclear (33). Stranges et al. (2010) (34) in their study on the association of high selenium with blood lipids observed a positive correlation between plasma selenium and total cholesterol and non-HDL cholesterol. This was one of the notable negative effects of elevated serum selenium.

Increased citrulline levels with elevated homocysteine is a novel finding in our study. The increase in the serum levels of homocysteine causes decreased nitric oxide (NO) bioavailability by inhibiting endothelial nitric oxide synthase (eNOS) induced by asymmetric dimethylarginine (ADMA) in hyperhomocysteinemia. Plasma levels of asymmetric dimethylarginine (ADMA) increase rapidly after acute methionine intake, which increases intracellular levels of S-adenosylmethionine (SAM). This accelerates the production of ADMA by inducing SAM-dependent protein arginine methyltransferase activity (PRMT) (35). Homocysteine synthesized by hydrolysis of SAH remethylates into methionine (Figure 3), where the increase in the intake of methionine and high serum levels of homocysteine result in the conversion of arginine residues in the dietary protein into methylated protein. This results in the increase of ADMA by proteolysis of methylated protein, which inhibits eNOS resulting in the decrease of NO production driving endothelial dysfunction. Homocysteine may also elevate the serum ADMA alternatively by inducing endoplasmic reticulum stress and apoptosis, which leads to protein degradation resulting in methylarginine residues. Dimethylaminohydrolase (DDAH) is the major enzyme involved in the metabolism of ADMA, which produces citrulline and methylamine by hydrolysis of ADMA. The increase in serum levels of homocysteine or citrulline indicates NO-mediated endothelial dysfunction (36). The parallel increasing trend of homocysteine and citrulline (p<0.05) in the present study might be associated with ADMA-mediated citrulline synthesis.

Increased levels of homocysteine can be associated with toxicity during arthritis treatment with methotrexate. Initial homocysteine levels were predictive of toxicities such as gastrointestinal intolerance and elevations of liver enzymes (37). Methotrexate treatment causes increased serum levels of homocysteine

in rheumatoid arthritis and other diseases. However, toxicity-related discontinuation of methotrexate treatment was not associated with the change in homocysteine concentration. Methotrexate works by inhibiting an enzyme involved in the metabolism of folate, which can lead to an accumulation of homocysteine in the blood. One strategy to manage increased homocysteine levels in arthritis patients receiving methotrexate is to supplement with folic acid, which can help to lower homocysteine levels. Other strategies may include dietary changes, such as increasing intake of foods rich in folate and other B vitamins, or adding additional vitamin supplements (38, 39).

Additionally, in the present study, a diverse class of amino acids was evaluated for homocysteine-mediated cardiovascular disorders. Among the analyzed branched-chain amino acids (BCAA), isoleucine was found to have a considerable positive correlation with homocysteine. BCAAs have shown associations in the development of metabolic syndrome which involves increased levels of plasma homocysteine (40). A positive correlation was also observed between carnitine and serum homocysteine. Several studies have reported the clinical significance of carnitine when supplemented as Acetyl-L-Carnitine or Propionyl- L -Carnitine. However, being metabolites of exogenous amino acids both homocysteine and carnitine are independent indicators of abnormal amino acid metabolism (41). Hence the association of carnitine with homocysteine remains complicated and requires more research.

Homocysteine can be converted to methionine through a process called remethylation. This reaction is catalyzed by either methionine synthase or betainehomocysteine methyltransferase (Fig 3). The conversion of homocysteine to methionine requires vitamin B12 and folic acid as cofactors. A deficiency of these vitamins may lead to increased homocysteine levels. Homocysteine and its precursor methionine have been found to decrease reactive oxygen species (ROS) production in erythrocytes and inhibit erythrocytes' osmotic fragility increase and methemoglobin formation (42). Homocysteine was also demonstrated to enhance ferryl Hb reduction. Methionine is a precursor of homocysteine, and it can regulate metabolic processes, the innate immune system, and digestive functioning in mammals (43). The methionine-homocysteine cycle is a vital process that exists at a critical biochemical intersection in the methionine cycle. High blood levels of homocysteine signal a breakdown in this process, which can result in far-reaching biochemical and life consequences.

Gene mutations that could be responsible for the increased homocysteine levels could not be considered in the study. Apart from this limitation, the present study has several highlights such as understanding the complete nutrient profile including a diverse class of micronutrients - vitamins, minerals, and amino acids. The study also presents a clear statement on the association of vital cofactors involved in homocysteine metabolism. To the best of our knowledge, this is the first report that studied the association of a diverse class of micronutrients with serum homocysteine and reported several novel associations.

Conclusion

The present study confirms the association of various dietary nutrients with elevated levels of homocysteine. Apart from the conventional biomarkers vitamins B_9 and B_{12} , the current study signifies the association of other nutrients namely vitamin A, selenium, and amino acids asparagine, cystine, isoleucine, citrulline, and carnitine as vital biomarkers affecting the levels of homocysteine. The results motivate further research into the role of nutrition in modulating the levels of inflammatory markers such as homocysteine which is of vital importance in preventive care settings.

Competing Interests: Krishnamurthy, Jayaraman, Krishna, Wang, Bei, and Rajasekaran are employees of Vibrant Sciences LLC. Reddy, Song, and Rajasekaran are employees of Vibrant America LLC. Vibrant America is a commercial diagnostic lab that could benefit from increased testing of micronutrients and cardiovascular biomarkers.

Acknowledgments: We acknowledge Vibrant America LLC for supporting this research.

Availability of Data and Materials: To access supporting data, contact the corresponding author.

Authors' Contributions: Hari Krishnamurthy, Karthik Krishna, and Tianhao Wang performed the research. Hari Krishnamurthy, John J. Rajasekaran, Karenah Rajasekaran, and Vasanth Jayaraman designed the study. Qi Song, Kang Bei, and Swarnkumar Reddy analyzed the data. Hari Krishnamurthy and Swarnkumar Reddy wrote the article.

Institutional Review Board Statement: The study comprises retrospective analysis exempted by the Western Institutional Review Board.

References

- 1. Kreatsoulas C, Anand SS. The impact of social determinants on cardiovascular disease. Can J Cardiol. 2010 Aug-Sep;26 Suppl C:8C-13C. doi: 10.1016/s0828-282x(10)71075-8. PMID: 20847985; PMCID: PMC2949987.
- Rahman K. and Lowe G.M. Garlic and cardiovascular disease: a critical review. J Nutr 2006; 136(3); 736S-740S.
- Krishnamurthy H.K, Reddy S, Jayaraman V, et al. Effect of Micronutrients on Thyroid Parameters. J. Thyroid Res 2021.
- Freeman D.J, Norrie J, Caslake M.J, et al. C-reactive protein is an independent predictor of risk for the development of diabetes in the West of Scotland Coronary Prevention Study. Diabetes 2002; 51(5): 1596-1600.
- Kaul S, Zadeh A.A, Shah P.K. Homocysteine hypothesis for atherothrombotic cardiovascular disease: not validated. J. Am. Coll. Cardiol 2006; 48(5): 914-923.
- 6. Streck E.L, Vieira P.S, Wannmacher C, et al. In vitro effect of homocysteine on some parameters of oxidative stress in rat hippocampus. Metab. Brain Dis 200; 18(2): 147-154.
- 7. McCully K.S. Homocysteine and vascular disease. Nat. Med 1996; 2(4): 386-389.
- 8. Zhou J, Møller J, Danielsen C.C, et al. Dietary supplementation with methionine and homocysteine promotes early atherosclerosis but not plaque rupture in ApoE-deficient mice. Arterioscler Thromb Vasc Biol 2001; 21(9): 1470-1476.
- 9. Obeid R and Herrmann W. Homocysteine and lipids: S-adenosyl methionine as a key intermediate. FEBS letters 2009; 583(8): 1215-1225.
- 10. Ali A.K. Establishing a relationship between serum homocysteine levels and disease severity in adults with sickle cell anaemia in Lagos university teaching hospital (LUTH) 2015; Lagos, Nigeria. Faculty of Pathology.
- Langman L.J, Cole D.E. Homocysteine. Crit. Rev. Clin. Lab. Sci 1999; 36(4): 365-406.
- 12. Herrmann W, Schorr H, Purschwitz K, Rassoul F, Richter V. Total homocysteine, vitamin B12, and total antioxidant status in vegetarians. Clin chem 2001; 47(6): 1094-1101.
- 13. Fenech MF, Dreosti IE, Rinaldi JR. Folate, vitamin B12, homocysteine status and chromosome damage rate in lymphocytes of older men. Carcinogenesis. 1997; 18(7): 1329-36.

- Cagnacci A, Baldassari F, Rivolta G, Arangino S, Volpe A. Relation of homocysteine, folate, and vitamin B12 to bone mineral density of postmenopausal women. Bone 2003; 33(6): 956-959.
- Thiamin R. Dietary reference intakes for thiamin, riboflavin, niacin, vitamin B6, folate, vitamin B12, pantothenic acid, biotin, and choline. 1998; 6015.
- 16. Satyanarayana A, Balakrishna N, Pitla S, et al. Status of B-vitamins and homocysteine in diabetic retinopathy: association with vitamin-B12 deficiency and hyperhomocysteinemia. PloS one 2011; 6(11): 26747.
- Młynarczyk M, Falkowska M, Micun Z, et al. Diet, Oxidative Stress, and Blood Serum Nutrients in Various Types of Glaucoma: A Systematic Review. Nutrients. 2022 Mar 29;14(7):1421. doi: 10.3390/nu14071421.
- Krishnan Krishnamurthy H, Reddy S, Jayaraman V, Krishna K, E. Rajasekaran K, Wang T, et al. Association of Micronutrients and Prevalence of Antibodies in Hyperthyroidism [Internet]. Hyperthyroidism-Recent Updates. IntechOpen; 2023. Available from: http://dx.doi.org/10.5772/intechopen .109375.
- 19. Rush E.C, Katre P, Yajnik C.S. Vitamin B12: one carbon metabolism, fetal growth and programming for chronic disease. Eur. J. Clin. Nutr 2014; 68(1): 2-7.
- 20. Aggrey S.E, González-Cerón F, Rekaya R, Mercier, Y. Gene expression differences in the methionine remethylation and transsulphuration pathways under methionine restriction and recovery with D, L-methionine or D, L-HMTBA in meat-type chickens. J Anim Physiol Anim Nutr 2018; 102(1): 468-e475.
- 21. Krishnamurthy HK, Reddy S, Jayaraman V, et al. Preliminary Study on the Association of Serum Branched-Chain Amino Acids With Lipid and Hepatic Markers. Cardiol Res. 2023 Jun;14(3):167-175. doi: 10.14740/cr1454. Epub 2023 May 26.
- 22. Ingenbleek Y. The oxidative stress of hyperhomocysteinemia results from reduced bioavailability of sulfur-containing reductants. Clin. Chem 2011; 4(1).
- 23. Vazquez-Lorente H, Herrera-Quintana L, Molina-López J, Gamarra Y, Planells E. Effect of zinc supplementation on circulating concentrations of homocysteine, vitamin B12, and folate in a postmenopausal population. J. Trace Elem. Med. Biol 2022; 71: 126942.
- Ostrakhovitch E.A, Tabibzadeh S. Homocysteine and ageassociated disorders. Ageing Res. Rev 2019; 49: 144-164.
- Mattson M.P, Kruman I.I, Duan W. Folic acid and homocysteine in age-related disease. Ageing Res. Rev 2002; 1(1): 95-111.
- 26. Raslova K, Bederova A, Gašparovič J, Blažíček P. Effect of Diet and 677 C¬» T 5, 10-Methylenetetrahydrofolate Reductase Genotypes on Plasma Homocyst(e)ine. Physiol. Res 2000; 49: 651-658.
- Skovierova H, Vidomanova E, Mahmood S, et al. The molecular and cellular effect of homocysteine metabolism imbalance on human health. International journal of molecular sciences 2016; 17(10):1733.

- 28. Kataria N, Yadav P, Kumar R, et al. Effect of vitamin B6, B9, and B12 supplementation on homocysteine level and cardiovascular outcomes in stroke patients: a meta-analysis of randomized controlled trials. Cureus 2021; 13(5).
- 29. Lyon P, Strippoli V, Fang B, Cimmino L. B vitamins and one-carbon metabolism: implications in human health and disease. Nutrients 2020; 12(9): 2867.
- 30. Ganji V, Kafai MR. Frequent consumption of milk, yogurt, cold breakfast cereals, peppers, and cruciferous vegetables and intakes of dietary folate and riboflavin but not vitamins B-12 and B-6 are inversely associated with serum total homocysteine concentrations in the US population. Am J Clin Nutr. 2004 Dec;80(6):1500-7. doi: 10.1093/ajcn/80.6.1500.
- Olsen T, Vinknes K.J, Svingen G.F, et al. The risk association of plasma total homocysteine with acute myocardial infarction is modified by serum vitamin A. Eur. J. Prev. Cardiol 2008; 25(15): 1612-1620.
- Puddu P.E. Serum vitamin A, total homocysteine and acute myocardial infarction: is there a causal association? Eur. J. Prev. Cardiol 2018; 25(15): 1607-1611.
- 33. González S, Huerta J.M, Fernández S, Patterson A.M, Lasheras C. Homocysteine increases the risk of mortality in elderly individuals. Br. J. Nutr 2007; 97(6): 1138-1143.
- 34. Gonzalez S, Huerta J.M, Álvarez-Uría J, Fernandez S, Patterson A.M, Lasheras C. Serum selenium is associated with plasma homocysteine concentrations in elderly humans. J Nutr 2004; 134(7): 1736-1740.
- 35. Stranges S, Laclaustra M, Ji C, et al. Higher selenium status is associated with adverse blood lipid profile in British adults. J Nutr 2010; 140(1): 81-87.
- 36. Myles P.S, Chan M.T, Kaye D.M, et al. Effect of nitrous oxide anesthesia on plasma homocysteine and endothelial function. The Journal of the American Society of Anesthesiologists 2008; 109(4): 657-663.
- 37. Jin L, Caldwell R.B, Li-Masters T, Caldwell R.W. Homocysteine induces endothelial dysfunction via inhibition of arginine transport. J. Physiol. Pharmacol 2007; 58(2): 191.
- 38. Zhu J, Wang Z, Tao L, et al. MTHFR Gene Polymorphism Association With Psoriatic Arthritis Risk and the Efficacy and Hepatotoxicity of Methotrexate in Psoriasis. Front Med (Lausanne). 2022 Apr 11;9:869912. doi: 10.3389 /fmed.2022.869912.
- 39. Katsushima M, Minamino H, Shirakashi M, et al. High plasma homocysteine level is associated with increased prevalence of the non-remission state in rheumatoid arthritis: Findings from the KURAMA cohort. Mod Rheumatol. 2023 Aug 25;33(5):911-917. doi: 10.1093/mr/roac106.
- 40. Cherif RA, Yargui L, Slimani S, et al. Impact of chronic low dose methotrexate treatment in rheumatoid arthritis on biochemical parameters in a population of eastern Algeria: A cross-sectional study. Algerian Journal of Health Sciences 2023 Jun 4;1:46-57.
- 41. Ntzouvani A, Nomikos T, Panagiotakos D, et al. Amino acid profile and metabolic syndrome in a male Mediterranean population: a cross-sectional study. Nutr Metab Cardiovasc Dis 2017; 27(11): 1021-1030.

- 42. Kokot F, Chudek J, Łysiak-SzydŁowska W, et al. Levels of carnitine and homocysteine in plasma of long-term hemodialysis patients with chronic renal failure. Polskie Arch. Med. Wewnetrznej 2001; 106(6): 1131-1136.
- 43. Katturajan R, S V, Rasool M, Evan Prince S. Molecular toxicity of methotrexate in rheumatoid arthritis treatment: A novel perspective and therapeutic implications. Toxicology. 2021 Sep;461:152909. doi: 10.1016/j.tox.2021.152909. Epub 2021 Aug 25.
- 44. Miller AL. The methionine-homocysteine cycle and its effects on cognitive diseases. Altern Med Rev. 2003 Feb;8(1):7-19. PMID: 12611557.

Correspondence:

Received: 26 August 2022 Accepted: 15 June 2023

Hari Krishnan Krishnamurthy, Director of Biomedical

Engineering

Vibrant Sciences LLC 1021 Howard Avenue San Carlos, CA 94070 E-mail: hari@vibrantsci.com

Appendix

Supplementary files

Table S1. General characteristics of the all-study subjects according to the categories of serum homocysteine

	Characteristics	Quartiles of serum homocysteine levels					
		Total (n=336)	Q1 (n=84)	Q2 (n=84)	Q3 (n=85)	Q4 (n=83)	p
	Homocysteine	9.9±3.0 (4.1-28.9)	6.9±0.9 (4.1-8.0)	8.6±0.3 (8.0-9.3)	10.2±0.6 (9.3-11.3)	14.0±3.1 (11.3-28.9)	
	Age	48.7±15.0	41.9±15.0	48.6±14.1	50.1±14.7	54.3±13.6	<0.0001
	Vitamin B1	21.6±15.0	19.8±12.0	24.8±19.3	21.6±14.3	20.0±13.2	0.1113
nins	Vitamin B2	28.2±25.0	26.3±18.3	34.1±35.7	28.5±22.7	23.9±18.3	0.0531
ritan	Vitamin B3	19.1±88.6	17.6±7.0	19.0±7.3	21.4±11.9	18.3±6.9	0.0232
Water- soluble vitamins	Vitamin B5	108.1±88.6	117.6±85.0	103.9±82.4	113.9±100.1	96.9±85.5	0.4184
solul	Vitamin B6	17.0±18.5	15.3±12.5	17.7±13.0	18.7±17.3	16.3±27.4	0.6603
er-	Vitamin B9	13.7±4.1	15.3±3.3	14.4±4.2	13.7±3.5	11.1±4.2	<0.0001
Wat	Vitamin B12	735.6±329.4	866.8±326.0	811.0±355.7	659.6±300.5	604.6±261.5	<0.0001
	Vitamin C	0.5±0.3	0.5±0.4	0.4±0.2	0.5±0.2	0.4±0.3	0.6345
s,	Vitamin A	77.5±24.4	72.9±24.3	77.2±24.5	78.3±21.0	81.4±27.2	0.1556
umin	Vitamin D3	0.9±0.3	0.9±0.3	1.0±0.3	0.9±0.3	0.9±0.3	0.1804
Fat – soluble vitamins	Vitamin D 25 OH	47.9±23.0	48.5±21.1	47.9±22.5	51.2±26.5	43.9±21.4	0.2329
solu	Vitamin K1	1.6±1.4	1.5±1.1	1.8±1.8	1.5±1.0	1.6±1.4	0.5231
at –	Vitamin K2	0.7±1.2	0.8±1.6	0.6±0.7	0.8±1.6	0.6±0.8	0.5561
14	Vitamin E	13.4±4.7	13.7±5.4	13.4±4.4	13.9±4.8	12.8±4.2	0.4778
	Selenium	143.5±29.1	139.1±22.6	138.2±24.6	150.1±40.5	146.5±23.3	0.0179
	Sodium	141.6±3.4	141.0±3.0	141.8±3.2	141.9±3.4	141.8±4.0	0.2324
	Potassium	4.5±0.4	4.6±0.4	4.5±0.4	4.6±0.4	4.5±0.4	0.3445
	Calcium	9.7±0.4	9.6±0.4	9.7±0.4	9.6±0.4	9.7±0.5	0.4712
Minerals	Manganese	0.7±0.5	0.8±0.7	0.7±0.3	0.7±0.4	0.7±0.3	0.5526
Min	Zinc	0.7±0.2	0.7±0.2	0.7±0.2	0.7±0.2	0.7±0.1	0.3060
	Copper	1.1±0.3	1.1±0.3	1.1±0.4	1.0±0.3	1.0±0.3	0.8545
	Chromium	0.3±0.2	0.2±0.1	0.3±0.1	0.3±0.2	0.2±0.2	0.2969
	Iron	103.3±39.1	101.5±42.3	104. 9±35.6	108.4±43.2	98.2±34.5	0.3682
	Magnesium	2.2±0.2	2.2±0.2	2.2±0.2	2.2±0.2	2.2±0.2	0.0751
ial	Asparagine	53.8±12.0	52.4±11.9	53.7±13.3	54.1±12.0	55.1±10.5	0.5552
Non-essential AA	Serine	147.7±34.3	145.3±32.8	151.7±36.7	149.9±36.6	143.7±30.4	0.3894

	Characteristics	Quartiles of serum homocysteine levels					
		Total (n=336)	Q1 (n=84)	Q2 (n=84)	Q3 (n=85)	Q4 (n=83)	p
A	Isoleucine	69.1±20.7	63.7±15.2	70.3±21.5	69.3±22.6	73.1±21.8	0.0271
BCA	Valine	246.2±55.8	235.9±51.8	252.5±59.7	243.8±54.3	252.5±56.4	0.1604
B	Leucine	163.8±34.9	155.2±33.9	164.6±36.1	167.3±34.0	168.0±34.5	0.0634
<u> </u>	Cysteine	18.4±9.3	14.0±6.8	17.5±7.7	18.7±8.4	23.5±11.3	<0.0001
nally AA	Glutamine	504.5±81.8	494.9±87.0	501.8±78.8	511.5±78.9	509.7±82.6	0.5305
ditio	Citrulline	30.2±8.6	28.3±9.1	29.7±7.5	30.3±7.6	32.6±9.5	0.0121
Conditionally essential AA	Arginine	123.0±33.6	122.5±32.4	121.7±33.0	123.7±33.5	124.3±36.0	0.9596
	Coenzyme Q10	2.1±0.9	1.9±0.9	2.1±0.9	2.2±0.9	2.2±1.1	0.1563
	Choline	14.0±5.7	13.0±5.5	13.9±5.8	14.4±5.5	14.7±5.8	0.2055
	Inositol	37.1±10.1	36.1±10.4	38.6±9.9	37.4±10.0	36.3±10.1	0.3716
	Carnitine	27.2±7.2	25.9±7.5	27.1±6.7	27.8±6.9	28.0±7.8	0.2253
	MMA	0.3±0.2	0.3±0.1	0.3±0.1	0.3±0.2	0.3±0.3	0.0337